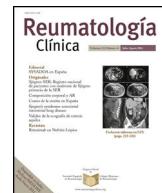




Sociedad Española
de Reumatología -
Colegio Mexicano
de Reumatología

Reumatología clínica

www.reumatologiaclinica.org



Case Report

Inclusion Body Myositis: A Late Diagnosis Case Report



Deysi Andrea Hernández-Rivero,^a Lisette Bazán-Rodríguez,^b
María del Pilar Cruz-Domínguez,^c Gabriela Medina,^d Ana Lilia Peralta Amaro,^e Olga Vera-Lastra^{e,*}

^a División Académica Multidisciplinaria de Comalcalco, Universidad Juárez Autónoma de Tabasco, Comalcalco, Tabasco, Mexico

^b Departamento de Enfermedades Neuromusculares, Hospital de Especialidades «Dr. Antonio Fraga Mouret», Centro Médico Nacional «La Raza», Mexico City, Mexico

^c Dirección de Investigación y Educación, Hospital de Especialidades «Dr. Antonio Fraga Mouret», Centro Médico Nacional «La Raza», Mexico City, Mexico

^d Unidad de Investigación Traslacional, Hospital de Especialidades «Dr. Antonio Fraga Mouret», Centro Médico Nacional «La Raza», Mexico City, Mexico

^e Departamento de Medicina Interna, Hospital de Especialidades «Dr. Antonio Fraga Mouret», Centro Médico Nacional «La Raza», Mexico City, Mexico

ARTICLE INFO

Article history:

Received 3 May 2024

Accepted 5 July 2024

Available online 28 October 2024

Keywords:

Inclusion body myositis
Idiopathic inflammatory myopathy
Muscle weakness
Creatinine kinase
Rimmed vacuoles

ABSTRACT

Inclusion body myositis is a idiopathic inflammatory myopathy characterized by muscle weakness and dysphagia, with muscle biopsy showing inflammation and rimmed vacuoles. We present the case of a patient who was diagnosed with polymyositis but due to lack of response to treatment, a new biopsy revealed inclusion body myositis.

© 2024 Published by Elsevier España, S.L.U.

Palabras clave:

Miositis por cuerpos de inclusión
Miopatía inflamatoria idiopática
Debilidad muscular
Disfagia
Vacuolas ribeteadas

Miositis por cuerpos de inclusión: informe de un caso de diagnóstico tardío

RESUMEN

La miositis por cuerpos de inclusión es una miopatía inflamatoria idiopática caracterizada por debilidad muscular, disfagia y biopsia muscular con inflamación y vacuolas ribeteadas. Presentamos el caso de una paciente a la que se le diagnosticó polimiositis pero que por falta de respuesta al tratamiento se le realizó una nueva biopsia que reveló miositis por cuerpos de inclusión.

© 2024 Publicado por Elsevier España, S.L.U.

Introduction

Inclusion body myositis (IBM) is an immune-mediated disease affecting muscles and internal organs.¹ Prevalence rates vary from 24.8 to 45.6 per million.² It affects individuals over the age of 50³ and debuts with musculoskeletal damage to the volar aspect of the forearm, flexors of the fingers and quadriceps, resulting in asymmetrical muscle weakness and inclusion bodies on biopsy.⁴ Biochemically, they are accompanied by elevated creatine phosphokinase (CPK), the presence of anti-cytosolic 50-nucleotidase CN1A antibody in up to 30% of cases, which, though non-specific, are associated with increased severity and mortality, and biopsy

reveals an endomysial inflammatory exudate with infiltration of inflammatory cells in non-necrotic muscle fibres and vacuoles rimmed by membranous cytoplasmic material, atrophic fibres, and intra- or extravacuolar congophilic inclusions.³

We report the case of a female patient diagnosed with polymyositis (PM) on the basis of elevated CPK, positive AC-1 antibodies, electromyography (EMG) with myopathic pattern, and muscle biopsy with inflammatory infiltrate, with poor response to steroid treatment. A new biopsy was therefore carried out that demonstrated IBM.

Case Report

An 82-year-old female presented difficulty climbing stairs and swallowing, diminished muscle strength in the shoulder girdle and pelvis, elevated CPK levels, and was positive for AC-1 antibod-

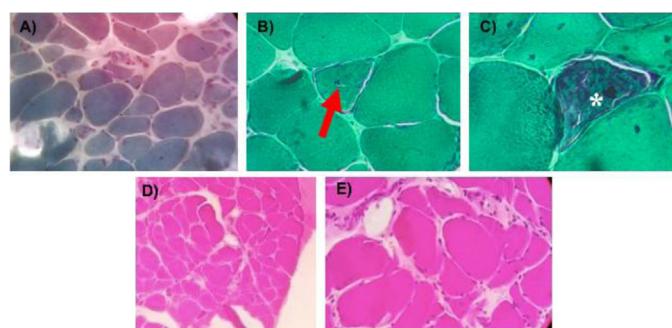
* Corresponding author.

E-mail address: olgavera62@yahoo.com.mx (O. Vera-Lastra).

Table 1

Baseline laboratory values of the patient with inclusion body myositis.

| Baseline lab | Results | Reference value |
|------------------------|-----------|-----------------|
| <i>Blood count</i> | | |
| Haemoglobin | 13.2 g/dL | 12–18 g/dL |
| Haematocrit | 40.1% | 37%–52% |
| Leukocytes | 3.4 K/UL | 4.5–10 K/UL |
| Platelets | 252 K/UL | 150–250 K/UL |
| <i>Blood chemistry</i> | | |
| Creatinine kinase | 936 UI/L | 24–170 UI/L |
| Lactate dehydrogenase | 867 UI/L | 123–245 UI/L |
| Alanine transferase | 61 U/L | 13–40 U/L |
| Aspartate transferase | 59 U/L | 9–36 U/L |
| <i>Antibodies</i> | | |
| AC-1 | 1:1280 | <1:80 |
| Anti-Jo | 1:98 | <20 |

**Figure 1.** Muscle biopsy from this case.

Modified trichrome Gomori (TMG) staining. Close-up 200× TMG, staining demonstrated muscle fibres of varying size, ballooning, aggregates of inflammatory cells with focal invasion of some muscle fibres, and substantial proliferation of connective tissue (A). Close-up 400× TMG, with obvious nuclear centralisation (arrow) (B). Close-up 400× TMG Vacuoles rimmed with granular material and filaments (star) classically observed in this pathology (C). Haematoxylin and eosin (H&E) staining. Close-up 100×, panoramic view of the biopsy displaying a major deviation from the general architecture, with enlarged spaces and substitution by connective tissue, fibre morphology with bulging and inflammatory infiltrate (D). Close-up at 400× (H&E) illustrates the inflammatory infiltrate surrounding the muscle fibre in greater detail (E), as well as abundant connective tissue.

ies at the age of 66. Anti-Jo1 immunospecificity was subsequently assessed and was negative (Table 1). An EMG examination was conducted that evidenced myopathic pattern and a biopsy of the left deltoid muscle was performed with the patient's informed consent and endomysial lymphocytic inflammatory infiltrate was noted. The woman was diagnosed with MP and treatment was instituted with prednisone and methotrexate, with a subsequent change to mycophenolate mofetil.

Her disease progressed and she required the use of a cane and wheelchair; she experienced difficulty in extending the fingers of the left hand, atrophy of the long flexor muscles of the fingers, predominantly the left one, and quadriceps. Muscle testing (MMT8/MMT26 manual muscle assessment procedures) yielded a score of 116 of 150 and 194 of 260, respectively, at the expense of distal muscle involvement. The new muscle biopsy indicated the presence of an inflammatory infiltrate with rimmed vacuoles and IBM was diagnosed (Fig. 1).

Discussion

This case initially featured symmetrical proximal muscle weakness, dysphagia, and elevated CPK, as well as an endomysial lymphocytic infiltrate, characteristic of PM.⁵ Nonetheless, recurrence of elevated CPK values, increased proximal muscle weakness and distal involvement, asymmetric atrophy of the hand and quadriceps muscles, deterioration of the patient's dysphagia, and

her poor response to treatment led to IBM2 being suspected. Alamri et al. reported that 14% of 367 patients with IBM had an atypical onset and of them, 6% displayed proximal arm weakness at the time of onset.⁶ In a clinical case series of IBM with no rimmed vacuoles on biopsy, diagnosis was delayed for up to eight years following symptom debut⁷; however, this feature may be absent in 20% of all cases.⁸ The condition that imitates IBM most closely is PM, inasmuch as its serology is similar and antinuclear antibodies (ANA) are positive in 15–19% in IBM and 60% in PM.⁹ A retrospective study comparing several subtypes of inflammatory myopathies failed exhibit any significant difference between PM and IBM with respect to follow up, disease duration, clinical characteristics, and CPK values.¹⁰ In fact, the presence of PM has currently been called into question, as most individuals diagnosed with PM are re-classified as suffering from anti-synthetase syndrome having no rash, immune-mediated necrotising myopathy, or IBM with or without rimmed vacuoles ribeteadas.^{3,11}

The clinical course in this case evidences the challenge posed by the differential diagnosis between PM and IBM, which entails a delay in establishing the diagnosis.

Conclusion

The clinical evolution with asymmetrical muscle involvement and poor response to immunosuppressants made it possible to reformulate the diagnosis as IBM, with a slowly progressive course as well as histopathological changes in muscle biopsies. Treatment with corticosteroids and immunosuppressants failed to stem the progression of the disease.

Funding

No external funding was necessary. The resources of the Hospital de Especialidades Centro Médico Nacional, La Raza were used.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.reumae.2024.10.002>.

Declaration of competing interest

The authors have no conflict of interests to declare.

References

- Schmidt J. Current classification and management of inflammatory myopathies. *J Neuromuscul Dis.* 2018;5:109–29.
- Naddaf E, Barohn RJ, Dimachkie MM. Inclusion body myositis: update on pathogenesis and treatment. *Neurotherapeutics.* 2018;15:995–1005.
- Selva-O'Callaghan A, Pinal-Fernandez I, Trallero-Araguás E, Milisenda JC, Grau-Junyent JM, Mammen AL. Classification and management of adult inflammatory myopathies. *Lancet Neurol.* 2018;17:816–28.
- Chinoy H, Lilleker JB. Pitfalls in the diagnosis of myositis. *Best Pract Res Clin Rheumatol.* 2020;34:101486.
- Yang SH, Chang C, Lian ZX. Polymyositis and dermatomyositis – challenges in diagnosis and management. *J Transl Autoimmun.* 2019;2:100018.
- Alamri M, Pinto MV, Naddaf E. Atypical presentations of inclusion body myositis: clinical characteristics and long-term outcomes. *Muscle Nerve.* 2022;66:686–93.
- Vivekanandam V, Bugiardini E, Merve A, Parton M, Morrow JM, Hanna MG, et al. Differential diagnoses of inclusion body myositis. *Neurol Clin.* 2020;38:697–710.
- Greenberg SA. Inclusion body myositis: clinical features and pathogenesis. *Nat Rev Rheumatol.* 2019;15:257–72.
- Balakrishnan A, Aggarwal R, Agarwal V, Gupta L. Inclusion body myositis in the rheumatology clinic. *Int J Rheum Dis.* 2020;00:1–10.
- Chinniah KJ, Mody GM. The spectrum of idiopathic inflammatory myopathies in South Africa. *Clin Rheumatol.* 2020;40:1437–46.
- Tanboon J, Nishino I. Classification of idiopathic inflammatory myopathies: pathology perspectives. *Curr Opin Neurol.* 2019;32(5):704–14.