Fig. 2. Positive patch tests 72 h after exposure to methylisothiazolinone (top) and ultrasound gel (bottom).

References

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Pulmonary Cyst Associated With Primary Sjögren’s Syndrome

Quistes pulmonares asociados a síndrome de Sjögren primario

To the Editor,

More than 50% of the patients with primary Sjögren’s syndrome (pSS) have extraglandular manifestations, and pulmonary manifestations are observed in 10% of this patient population.1 In 50% of the patients, the involvement is subclinical; however, symptomatic lung disease occurs in 10%. Interstitial lung disease is the most common disorder.2–4 Cystic lung disease is reported much less frequently. This complication is characterized by foci of reduced lung density that have well-defined, thin walls (wall thickness less than 4 mm) and a diameter at their largest point of 1 cm.5 We report the cases of 3 patients with cystic lung disease associated with pSS. All of them met the 2002 American-European classification criteria for the diagnosis of Sjögren’s syndrome (SS).5

The 3 pSS patients were diagnosed, by means of high-resolution computed tomography (HRCT) of the thorax, as having associated cystic lung disease, together with centrilobular pulmonary emphysema, predominantly in the upper lobes in 2 of the 3 patients. There was no evidence of association with other lung diseases. We did not perform a histological examination, a study that had been described in other reports published up to that time.

Patient no. 1

The first patient was a 74-year-old woman with an 18-year history of pSS. She presented with extraglandular manifestations, such as the vasculitis, polyarteritis nodosa. The most notable laboratory findings were hypergammaglobulinemia and anti-Ro and anti-La antibodies. She was being treated with oral azathioprine at a dose of 50 mg/day. During follow-up she developed functional dyspnea grade II. The results of respiratory function tests were normal, and HRCT of the thorax revealed centrilobular pulmonary emphysema, predominantly in upper lobes, and images of thin-walled cysts predominantly in lower lobes (Fig. 1a). The same treatment was maintained, and there were no changes in the pulmonary lesions in subsequent visits.

Patient no. 2

This patient was a 46-year-old woman with a 20-year history of pSS and diagnosed with IgA kappa myeloma. Her extraglandular manifestations were anemia and leukocytoclastic vasculitis, as well as polyarteritis and parotid gland enlargement. She was being treated with hydroxychloroquine (HCQ) 200 mg/day, oral pilocarpine 15 mg/day and rituximab every 6 months. Laboratory tests revealed elevated acute-phase reactant levels, anemia and lymphopenia, as well as hypergammaglobulinemia with an elevated IgG level and a monoclonal IgA kappa component. Tests for rheumatoid factor (RF), as well as anti-Ro and anti-La antibodies, were positive. During follow-up, she developed dyspnea on heavy exertion. Respiratory function tests revealed a mild restrictive ventilatory defect with a slightly reduced diffusing capacity of the lung for carbon monoxide (DLCO), and HRCT of the thorax

showed bilateral cystic lung lesions—some with septa in their interior, traction bronchiectasis in both lower lobes, areas of centrilobular and paraseptal emphysema and zones of fibrotic tissue; and (c) image showing multiple bilateral thin-walled pulmonary cysts.

**Patient no. 3**

The third patient was a 78-year-old woman with a 15-year history of pSS who was also diagnosed with cutaneous amyloidosis. The manifestations were predominantly glandular (xerostomia and xerophthalmia), and there was no extraglandular involvement. She was receiving oral HCQ 200 mg/day. The laboratory tests revealed elevated acute-phase reactant levels, polyclonal hypergammaglobulinemia with an elevated IgG level, and positive tests for RF and anti-Ro and anti-La antibodies. During follow-up, she reported dyspnea on moderate exertion and, thus, underwent respiratory function tests, which revealed a mild restrictive ventilatory defect, with a severe reduction in DLCO. Thoracic HRCT (Fig. 1c) showed multiple bilateral thin-walled pulmonary cysts, with no evidence of nodules or changes in the air space. The findings were the same in subsequent visits.

Cystic lung disease is an uncommon condition that is rarely associated with an autoimmune inflammatory disease. The prevalence of pulmonary cysts in pSS ranges between 12% and 30% of the overall group of pulmonary manifestations. They can be found alone or in association with areas of ground glass attenuation.\(^7\) Two hypotheses have been proposed for the formation of these pulmonary cysts, both of which involve the infiltration of immunoglobulin-producing lymphocytes: a valve mechanism that causes cystic changes in the dilated alveolar region as a result of chronic inflammation and lymphoplasmocytic infiltration in peripheral regions; and the destruction of the alveolar wall, in which the direct lymphoplasmocytic infiltration of the alveoli destroys the alveolar structures and creates cystic areas. The most probable scenario involves both mechanisms.\(^7,8,10\)

There have been few reports of pulmonary cysts associated with pSS, and the majority of the cases are found to be concomitant with other diseases, such as lymphoid interstitial pneumonitis, lymphoproliferative processes and pulmonary amyloidosis.\(^11,12\)

One common characteristic finding is the high erythrocyte sedimentation rate with hypergammaglobulinemia observed in all 3 cases, a direct reflection of the lymphocyte hyperactivity associated with the disease. Compared with the cases in the literature, our patients do not differ with respect to the characteristic radiological findings,\(^10\) although there have been reports of a significant association between seropositivity for anti-SSB/La and clonal lymphoproliferative disorders that we have not identified in our patients.\(^7\)

**References**

Adverse Effects of Immunosuppressive Therapy in Rheumatic Patients: Non-tuberculous Mycobacterial Infection

Efectos adversos de terapia inmunosupresora en paciente reumatólogo: infección por micobacterias no tuberculosas

To the Editor,

Mycobacterium chelonae is a rapidly growing mycobacterium that causes skin and soft tissue infections after trauma in immunocompetent individuals and in immunocompromised hosts with disseminated disease, especially in patients with rheumatic diseases receiving immunosuppressive therapy.1-3

We report the case of a 29-year-old woman whose medical record included systemic lupus erythematosus, sarcoidosis and a 4-year history of sickle cell anemia plus β-thalassemia. At the time of writing, she was being treated with rituximab (she had received 2 cycles of 1 g each), azathioprine 100 mg/day, methylprednisolone 16 mg/day and hydroxyurea 1 g/day. She was admitted to a quaternary care center with a 3-month history of pain in her right hip. Drug treatment with a variety of analgesics and multiple local injections in right hip and gluteus resulted in a partial improvement of her symptoms. One month later, the clinical symptoms returned and she presented with an abscess in right gluteus, with local inflammatory changes. Physical examination revealed the presence of a painful nodule. She underwent surgical drainage of the abscess, and cultures for the usual bacteria, fungi and mycobacteria (auramine–rhodamine stain and culture procedures; (b) the presence of painful subcutaneous nodules and abscesses at the site of the lesion; and (c) a poor or inadequate response to the antibiotic therapy received.6

The diagnosis starts with the direct observation of mycobacteria in the aspirate of secretions or in the tissue obtained, using Ziehl–Neelsen or auramine–rhodamine stain, plus the performance of special cultures for the diagnosis. The classical Ziehl–Neelsen technique and the fluorescence technique with auramine–rhodamine are equally effective for the diagnosis, but around 30% of the rapidly growing mycobacteria can exhibit negative fluorescence with the auramine–rhodamine technique. Thus, when infection by rapidly growing mycobacteria is suspected, the procedure of choice is the Ziehl–Neelsen stain.7

Bacteriological culture enables us to enhance the sensitivity of the diagnosis; even if the result of direct observation is negative, the disease is not ruled out. Therefore, sampling should be carried out for DNA amplification using the PCR technique, to obtain the patterns of sensitivity to first-line and second-line drugs that serve as a guide to the most adequate and effective treatment.7

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